**Make 2x Wash Buffer (**There is ~50mL with our reagents, don’t have to make every time)

10mM Tris-HCl (ph 7.5)

1mM EDTA

2M NaCl

Wash Dynabeads

* vortex beads in vial for >30 sec
* Transfer desired volume to eppi tube (20µL)
* Add 1mL 1x wash buffer and resuspend (pipette to mix)
* Place on magnet 1 minute and discard supernatant
* Remove tube from magnet and resuspend beads in original volume of 1x buffer (20µL)
* Place on magnet 1 minute and discard supernatant
* Remove tube from magnet and resuspend beads in original volume of 1x buffer (20µL)
* Place on magnet 1 minute and discard supernatant

**Immobilize Nucleic Acids**

* resuspend beads in 2x wash buffer to twice original volume (40µL)
* add an equal volume of DNA (40µL)
* Incubate at room temp 15 min on shaker plate (1200 rpm is the slowest so that beads don’t accumulate on the bottom)
* Place on magnet 2 minutes, discard supernatant
* Wash 2 times with 1x buffer
* Resuspend in 40µL TE buffer